

RESEARCH PAPER

Effectiveness test of microfilter mask based on straw waste cellulose fiber biopolymer

Hazi Rofiqoh^{a*}, Baiq Hizanatul Ummah^b, Imam Galih Hanafi^a, Baiq Nely Budhiartini^a, Murniati^b Chang-Nan Chen^c

- [a] Department of Chemistry, Faculty of Teacher Training and Education, University of Mataram, Indonesia Majapahit Rd. No. 62, Mataram, West Nusa Tenggara, 83125, Indonesia
- [b] Department of Chemistry, Faculty of Mathematic and Natural Science, University of Mataram, Indonesia Majapahit Rd. No. 62, Mataram, West Nusa Tenggara, 83125, Indonesia
- [c] Department of Applied Chemistry, Chaoyang University of Technology, Taichung, Taiwan

E-mail: hazirofiqoh@gmail.com

DOI: 10.29303/aca.v8i1.172

Article info:	Abstract: Straw waste, which is often underutilized, contains abundant
Received 22/08/2023	cellulose that holds potential as a raw material for environmentally friendly filtration technologies. The conversion of agricultural waste into
Revised 27/08/2024	value-added products, such as mask microfilters, aligns with the increasing demand for sustainable materials, particularly in the context
Accepted 21/12/2024	of public health protection. Harnessing cellulose from straw offers a promising alternative to synthetic fibers commonly used in mask
Available online 30/05/2025	production, while simultaneously addressing waste management challenges. This study aims to determine the characterization of microfilter masks from straw waste and the effectiveness of straw microfilters used in cloth masks. Cellulose from straw waste can be used as a mask microfilter by going through several stages, such as: preparation, extraction, delignification, bleaching, characterization, and making mask microfilters and effectiveness tests. The characterization of structure, morphology and topology from the straw waste cellulose has not been fully degraded from lignin as shown from the aromatic C = C stretching vibrations on lignin at wave numbers 1638.34 cm ⁻¹ , 1648 cm ⁻¹ , and 1583.56 cm ⁻¹ of FTIR also SEM tools at 3600 times magnification. The effectiveness of the microfilter mask is 94% and 91% as evidenced by each bacterial filtration test and the decreasing data of the dust filtration test.

Keywords: Microfilter Mask, Cellulose, Straw Waste

Citation: Rofiqoh, H., Ummah, B. H., Hanafi, I. G., Budhiartini, B. N., Murniati, & Chen, C. N., 2025. Effectiveness Test of Microfilter Mask Based on Straw Waste Cellulose Fiber Biopolymer. *Acta Chimica Asiana*, 8(1), 606–611. https://doi.org/10.29303/aca.v8i1.172

INTRODUCTION

1. Introduction

The increasing awareness of environmental sustainability and public health protection has driven the search for alternative, environmentally friendly materials for various applications, including air filtration technology [1,2]. Agricultural waste, especially straw, is one of the abundant yet underutilized biomass sources [3]. Straw is mainly composed of cellulose, hemicellulose, and lignin, with cellulose being a particularly valuable component due to its

structural strength, renewability, and biodegradability [4]. Converting straw waste into value-added products not only promotes sustainable waste management but also supports the development of environmentally friendly alternatives to synthetic materials [5].

The use of cellulose extracted from straw waste as a microfilter in cloth masks is a promising innovation. Traditional mask filters usually rely on synthetic polymers, which are often non-biodegradable and contribute to environmental pollution after disposal [6]. In contrast, cellulose-based filters offer a biodegradable solution without compromising filtration performance, thus aligning with global efforts to reduce plastic waste and promote green technologies [7].

The production of cellulose-based microfilters involves several critical steps. including preparation, extraction, delignification, [8]. bleaching. and characterization Α comprehensive understanding of the chemical structure, morphology, and topology of the extracted cellulose is essential to optimize filtration efficiency. Characterization techniques such as Fourier transform infrared spectroscopy (FTIR) and scanning electron microscopy (SEM) are used to assess the lignin removal efficacy and evaluate the physical structure of cellulose fibers [9,10].

In addition to its abundance, the utilization of straw waste presents a more scalable and accessible solution compared to other agricultural residues such as pineapple leaves [11]. While pineapple leaf fibers have demonstrated promising filtration performance, their availability is relatively limited in certain regions of West Nusa Tenggara [12]. In contrast, rice straw is generated in much larger quantities as a byproduct of the dominant agricultural activity in Lombok [13]. Leveraging this locally available resource could not only reduce environmental pollution caused by unprocessed straw disposal but also provide a sustainable raw material for the production of biodegradable microfilter masks [14]. Furthermore, the high cellulose content in straw, coupled with the substantial silica composition, offers structural advantages for microfilter applications by enhancing mechanical stability and filtration capacity [15]. Therefore, utilizing straw waste for the development of cloth mask microfilters represents an innovative and regionally appropriate strategy to improve community-level protection while simultaneously health addressing agricultural waste management challenges [16].

MATERIALS AND METHODS

Straw waste was obtained from a farm located in Central Lombok, West Nusa Tenggara, Indonesia. Distilled water (H₂O), hydrogen peroxide (H₂O₂) (5%), sodium hydroxide (NaOH) (20%), toluene (C₆H₅CH₃) (85%), ethanol (C₂H₅OH) (96%), phosphate buffered saline (PGBP) diluent, and sterile liquid *Plate Count Agar* (PCA). This research also involved instruments such as Microbial Air Sampler (MAS 100-NT) (Merck) to microbial monitoring of ambient air in controlled environments, colony counter and inhibitors (35°C) to bacterial filtration test, and analytical balance to dust filtration test, also Fourier Transform-Infra Red (FTIR) (Parkin Elmer), Scanning Electron Microscope (SEM).

Cellulose Isolation

To obtain cellulose from straw waste, sample preparation, extraction, delignification, and bleaching were carried out.

In the sample preparation, the wet straw waste was cleaned and then dried for ± 3 days in the sun until dry. The dried straw waste was then cut into ± 2 -4 cm pieces and then smashed and filtered with a 40 mesh sieve. Extraction stage: Straw powder was reacted with toluene: ethanol 2:1 (v/v) then stirred until mixed then macerated for 8-24 hours. The mixture was filtered and the precipitate was washed with distilled water until the pH of the filtrate was neutral and then dried. Delignification: The residue was added to a 20% NaOH solution, in a ratio of 1:10 then the mixture was heated at 80 °C and stirred using a *magnetic stirrer* for 2 hours.

Afterwards, the mixture was filtered and the precipitate was washed with distilled water about 500 mL until the pH of the filtrate was neutral. The residue was dried at 50 °C to constant weight. Then the *bleaching* process was carried out by adding 5% H₂O₂ solution, pH 12 in a ratio of 1:10, then heated in a water bath at 70 °C and stirred using a magnetic stirrer which was kept constant for 3 hours. The mixture was treated with the same methode until the pH was neutral.

Cellulose Characterization

Cellulose produced from straw waste was characterized by FTIR to determine and analyze the vibrations of its functional groups, while SEM analysis was used to determine its morphology and topology.

Microfilter Making from Straw Cellulose

Cellulose is put into a tub containing clean water then put into a 40mesh sieve and then shaken until flat. Cellulose that has been flat and printed on the sieve is dried using a hair dryer until it is separated from the sieve and then dried in the sun.

Microfilter Mask Effectiveness Test

Bacterial filtration test:

The sterile microfilter was placed in MASS 100-NT at the top and bottom of the bag and then turned on MASS 100-NT with a suction speed of

500 m/s for 2 minutes. The microfilter was then inserted into the diluent and then homogenized. Serial dilutions were carried out with dilutions of 10^{-1} , 10^{-2} , 10^{-3} , and 10^{-4} using PGBP (phosphate buffered saline diluent) then poured sterile liquid count agar plate (sterile liquid PCA) while leveled. The plate was incubated at 35.5 °C for 2x24 hours then read and counted the number of bacterial colonies.

Dust filtration test. Sterilized microfilters were weighed with an analytical balance and then placed in a *Portable high-air sampler* for 10 minutes with a control speed of 70 m/s. After 10 minutes the sample was weighed with an analytical balance to determine the weight of dust obtained before and after filtration.

RESULTS AND DISCUSSION

X-Ray Diffraction (XRD)

Cellulose Isolation of Straw Waste

Straw waste that has gone through the preparation and extraction produces straw powder with a yield of 50.2 grams. The last stage of delignification and bleaching produces white cellulose. The average cellulose yield produced was 14.2 grams (Figure 1). The success of the straw waste cellulose isolation stage can be seen from the white cellulose [17].

The spectrum of straw cellulose shows a wide absorption at wave number 3445.84 cm⁻¹ which indicates the stretching vibration of a hydroxyl group (O-H) of cellulose [18]. The absorption at wave number 2905.34 cm⁻¹ indicates the stretching vibration of the aliphatic C-H group of cellulose. Wave numbers 1638.34 cm⁻¹, 1648 cm⁻¹, and 1583.56 cm⁻¹ show aromatic C=C stretching vibrations on lignin, meaning there is lignin in straw cellulose [19]. The absorption region of 1423.49 cm identifies the presence of CH₂ bending and bending absorption of C-H groups seen at wave numbers 1376.35 and 1316.52 cm⁻¹ [20]. The wavenumber 1159.49-1027.94 cm^{-1} is the area of C-O-C vibrations [21].

FTIR Characterization

Analysis of functional groups using FTIR characterization is carried out to determine the functional groups contained in the straw cellulose produced based on the wavenumber where a peak appears. The results of FTIR spectroscopic characterization show data on the

structure of the compounds contained in straw cellulose (Figure 2).



Figure 1. Cellulose powder

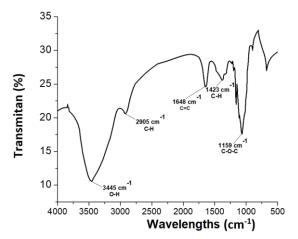


Figure 2. FTIR spectrum of straw cellulose

SEM Characterization

Morphological and topological analysis with SEM characterization was carried out to determine the surface morphology and see the topology on the surface of the straw cellulose produced with various magnifications. The results of cellulose powder characterization using SEM with 2000-4000x magnification obtained the size of straw waste cellulose around 29.2 µm -74.6 µm.

Figure 3 shows the morphology of cellulose with various magnifications on the SEM tool. The morphology of cellulose at 3600 times magnification is in the form of compact chunks with small cavities indicating that the lignin content has not been degraded. The 9200x magnification shows that the fibrils are still attached and some have not been defibrillated, indicating that the alkaline treatment and delignification procedures did not completely disrupt the cellulose structure and remove amorphous areas [22].

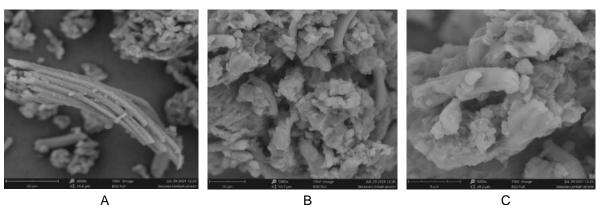


Figure 3. Morphology and topology of cellulose by SEM at magnifications of 3600x; b) 5000x; and, c) 9200x.

Mask microfilter

The mask microfilter of straw waste cellulose is white fibrous with a length of 14 cm, and a width of 9 cm. The weight of this straw waste microfilter is 1 gram which is biodegradable, so it does not cause environmental pollution. Microfilter masks made from cellulose are nontoxic based on a study of feeding rats with cellulose isolated through oral radicals [23]. In addition, cellulose shows a low toxicity impact [24].



Figure 4. Cellulose fiber microfilter

Mask Microfilter Effectiveness Test

Bacterial filtration test

Based on Table 1. in the bacterial filtration test using 3 controls, the number of bacterial colonies produced is 0, meaning that the three controls are not contaminated with bacteria. In Table 2 the number of bacterial colonies in the three samples decreased after being filtered with the microfilter.

Data in Table 2. shows the percentage of bacterial filtration obtained from the comparison of the difference in bacterial colonies before and after treatment with the number of previous colonies in samples I and II is 92% and 98%. Then, the percentage of bacterial filtration test results after the microfilter is used is obtained at 94%.

Table 1. Bacterial Filtration Res	ults with Control
-----------------------------------	-------------------

No	Control		Number of Bacterial Colonies
1	Deluent 9 mL		0
2	Deluent 90 mL		0
3	Deluent and	sterile	0
	microfilter		

Table 2. Bacterial Filtration Test Results onMicrofilters

<u> </u>	Treatment	Number of Bacterial Colonies				Bacterial Colony	
S		10- 1	10- 2	10- 3	10- 4	Count Results	
I	Before	16	1	0	0	130	
	After	1	0	0	0	10	
П	Before	1	2	2	0	736.67	
	After	1	0	0	0	10	

S = sample

Dust filtration test

Table 3 shows the comparison data before and after dust filtration. From the filtration result data after using the microfilter, there was a decrease in dust weight by 91%. Based on the quality standards set in Decree of the minister of environment (Kep-Men LH) No. 13 of 1995 related to the TSP (dust) parameter, the weight of dust that can be inhaled by humans is 35%. So, when viewed from the standard data, this straw waste cellulose fiber microfilter has high effectiveness because the dust filtration value produced by the microfilter is greater than 35%

Table 3. Dust Filtration Test Results						
Sample	Microfilter code	Mass I (g)	Mass II (g)	Mass of dust obtained (g)	A-B	
	А	0.05640	0.05687	0.00047	0.00043	
I	В	0.06202	0.06206	0.00004		
П	А	0.07571	0.07604	0.00033	0.00010	
	В	0.06785	0.06808	0.00023	0.00010	

CONCLUSION

Based on the research that has been done, it can be concluded that the cellulose obtained from straw waste is 14.7% per 100 grams of straw. Microfilters from straw waste cellulose fibers that have been tested for effectiveness with comparative data before and after filtration produce a decrease in the number of bacterial colonies by 94% and a decrease in dust weight by 91%.

ACKNOWLEDGEMENTS

The authors would like to express their gratitude and respect to Murniati S.Pd M.Si., Dr. Saprizal Hadisaputra, M.Sc., Early Sinthia Dewi S.T., M.Pd., and their support and direction in this research until completion.

REFERENCES

- [1] Owens, L. P., & Hubbe, M. A. (2023). Performance Factors for Filtration of Air Using Cellulosic Fiber-based Media: A Review. BioResources, 18(1), 2440-2519.
- [2] Siregar, S. Н., Rahmadini, S., Hasmalina, N., Rizki, R. A., & Eri, K. (2023). Pulp synthesis using bamboo raw materials through unbleached and bleached processes. Acta Chimica Asiana, 6(1), 247-253.
 - [3] Chen, M., Ma, Y., Xu, Y., Chen, X., Zhang, X., & Lu, C. (2013). Isolation and characterization of cellulose fibers from rice straw and its application in modified polypropylene composites. Polymer-Plastics Technology and Engineering, 52(15), 1566-1573..
- Sun, X. F., Sun, R. C., Fowler, P., & Baird, M. S. (2003). Isolation and [4] characterization of cellulose obtained by a two-stage treatment with organosolv and cyanamide activated hydrogen peroxide from straw. Carbohydrate Polymers, 52(4), 437-443.

- [5] Lu, H., Wang, X., & Zhang, L. (2018). Cellulose, nanocellulose, and antimicrobial materials the for development of biodegradable and reusable face masks. Bioresources.
- Rainey, T. J., et al. (2020). Developing [6] of Mask Material Using Nanocellulose to Fight against COVID-19. Sugar Asia Magazine.
- [7] Wang, J., Liu, S., Yan, X., Jiang, Z., Zhou, Z., Liu, J., ... & Jiang, W. (2021). Biodegradable and reusable cellulosebased nanofiber membrane preparation for mask filter by electrospinning. Membranes, 12(1), 23..
- [8] Nasri-Nasrabadi, B., Behzad, T., & Bagheri, R. (2014). Extraction and characterization of rice straw cellulose nanofibers optimized by an chemomechanical method. Journal of Applied Polymer Science, 131(7).
- [9] Jadaun, S., Upadhyay, N., & Siddiqui, S. (2025). Isolation and characterization of cellulose nanofibers from rice straw using ultrasonication-assisted extraction technique coupled with high shear dispersion. Biomass Conversion and Biorefinery, 1-17.
- Lu, P., & Hsieh, Y. L. (2012). [10] Preparation and characterization of cellulose nanocrystals from rice straw. Carbohydrate Polymers, 87(1), 564-573.
- [11] Sun, X., Wu, Q., & Ren, S. (2021). Recent advances in biomass-derived materials for personal protective equipment. Materials Today Sustainability, 12, 100059.
- Putri, R. D., & Khairurrijal. (2020). [12] Utilization of pineapple fiber as naturalbased nanofiber mask filter material. IOP Conf. Ser.: Mater. Sci. Eng., 852, 012089.

- [13] Badan Pusat Statistik Nusa Tenggara Barat. (2021). Statistik Pertanian Provinsi Nusa Tenggara Barat 2021.
- [14] Owens, L. P., & Hubbe, M. A. (2023). Performance Factors for Filtration of Air Using Cellulosic Fiber-based Media: A Review. BioResources, 18(1), 2440– 2519.
- [15] Liu, L., et al. (2022). Development of silica–cellulose composite membranes for advanced filtration applications. Carbohydrate Polymers, 287, 119353.
- [16] Zhao, M., Liao, L., Xiao, W., Yu, X., Wang, H., Wang, Q., ... & Cui, Y. (2020). Household materials selection for homemade cloth face coverings and their filtration efficiency enhancement with triboelectric charging. Nano Letters, 20(7), 5544–5552.
- [17] Lu, P. & Hsieh, Y. L. (2012). Preparation and Characterization of Cellulose Nanocrystals from Rice Straw. *Carbohydr Polym*, 87 (1), 564-573.
- [18] Kumar, A., Y. S. Negi, V.Choudhary & N. K.Bhardwaj. (2014). Characterization of Cellulose Nanocrystals Produced by Acid-Hydrolysis from Sugarcane Bagasse as Agro-Waste. *Journal of Materials Physics and Chemistry, 2* (1): 1-8.
- [19] Fatriasari, W., W.Syafii, N. Wistara, K. Syamsul & B. Prasetya. (2016). Lignin And Cellulose Changes of Betung Bamboo (Dendrocalamus Asper) Pretreated Microwave Heating. International Journal on Advanced Science Engineering Information Technology, 6 (2), 186-195
- [20] Nomanbhay, S.M., R. Hussain & K.Palanisamy. (2013). Microwave-Assisted Alkaline Pretreatment and Microwave Assisted Enzymatic Saccharification of Oil Palm Empty Fruit Bunch Fiber for Enhanced Fermentable Sugar Yield. Journal of Sustainable Bioenergy Systems, 3 (1), 7-17.
- [21] Supraba, W., Juliantoni, Y., & Ananto, A. D. (2021). The effect of stirring speeds to the entrapment efficiency in a nanoparticles formulation of java plumâ€[™] s seed ethanol extract (syzygium cumini). Acta Chimica Asiana, 4(1), 197-103.
- [22] Jonoobi, M., Harun, J., Mathew, A.P., Hussein, M.Z., & Oksman, K. (2010). Preparation of Cellulose Nanofibers with

Hydrophobic Surface Characteristic, *Cellulose*, *17*(1), 299-307.

- [23] Lin, N. & Dufresne, A. (2014). Nanocellulose in Biomedicine: Current Status and Future Prospect. *European Polymer Journal*, 59 (2014), 302-325.
- [24] Harper, B. J., Clendaniel, A., Sinche, F., Way, D., Hughes, M., Schardt, J., Simonsen, J., Stefaniak A. B., & Harper, S. L. (2016). Impacts of Chemical Modification on the Toxicity of Diverse Nanocellulose Materials to Depeloving Zebrafish. *Cellulose*, 23 (3), 1763-1775.